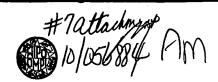


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(57) Abstract

Methods for isolating K+Hnov genes are provided. The K+Hnov nucleic acid compositions find use in identifying homologous or related proteins and the DNA sequences encoding such proteins; in producing compositions that modulate the expression or function of the protein; and in studying associated physiological pathways. In addition, modulation of the gene activity in vivo is used for prophylactic and therapeutic purposes, such as identification of cell type based on expression, and the like.

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HUMAN POTASSIUM CHANNEL GENES

INTRODUCTION

Background

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lon channels are multi-subunit, membrane bound proteins critical for maintenance of cellular homeostasis in nearly all cell types. Channels are involved in a myriad of processes including modulation of action potentials, regulation of cardiac myocyte excitability, heart rate, vascular tone, neuronal signaling, activation and proliferation of T-cells, and insulin secretion from pancreatic islet cells. In humans, ion channels comprise extended gene families with hundreds, or perhaps thousands, of both closely related and highly divergent family members. The majority of known channels regulate the passage of sodium (Na*), chloride (Cl*), calcium (Ca**) and potassium (K*) ions across the cellular membrane.

Given their importance in maintaining normal cellular physiology, it is not surprising that ion channels have been shown to play a role in heritable human disease. Indeed, ion channel defects are involved in predisposition to epilepsy, cardiac arrhythmia (long QT syndrome), hypertension (Bartter's syndrome), cystic fibrosis, (defects in the CFTR chloride channel), several skeletal muscle disorders (hyperkalemic periodic paralysis, paramyotonia congenita, episodic ataxia) and congenital neural deafness (Jervell-Lange-Nielson syndrome), amongst others.

The potassium channel gene family is believed to be the largest and most diverse ion channel family. K⁺ channels have critical roles in multiple cell types andpathways, and are the focus of significant investigation. Four human conditions, episodic ataxia with myokymia, long QT syndrome, epilepsy and Bartter's syndrome have been shown to be caused by defective K⁺ ion channels. As the K⁺ channel family is very diverse, and given that these proteins are critical components of virtually all cells, it is likely that abnormal K⁺ channels will be involved in the etiology of additional renal, cardiovascular and central nervous system disorders of interest to the medical and pharmaceutical community.

The K* channel superfamily can be broadly classified into groups, based upon the number of transmembrane domain (TMD) segments in the mature

protein. The minK (IsK) gene contains a single TMD, and although not a channel by itself, minK associates with different K⁺ channel subunits, such as KvLQT1 and HERG to modify the activity of these channels. The inward rectifying K+ channels (GIRK, IRK, CIR, ROMK) contain 2 TMD domains and a highly conserved pore domain. Twik-1 is a member of the newly emerging 4TMD K⁺ channel subset. Twik-like channels (leak channels) are involved in maintaining the steady-state K⁺ potentials across membranes and therefore the resting potential of the cell near the equilibrium potential for potassium (Duprat *et al.* (1997) <u>EMBO J</u> 16(17):5464-5471). These proteins are particularly intriguing targets for therapeutic regulation. The 6TMD, or Shaker-like channels, presently comprise the largest subset of known K⁺ channels. The slopoke (slo) related channels, or Ca⁺⁺ regulated channels apparently have either 10 TMD, or 6 TMD with 4 additional hydrophobic domains.

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Four transmembrane domain, tandem pore domain K+ channels (4T/2P channels) represent a new family of potassium selective ion channels involved in the control of background membrane conductances. In mammals, five channels fitting the 4T/2P architecture have been described: TWIK, TREK, TASK-1, TASK-2 and TRAAK. The 4T/2P channels all have distinct characteristics, but are all thought to be involved in maintaining the steady-state K* potentials across membranes and therefore the resting potential of the cell near the equilibrium potential for potassium (Duprat et al. (1997) EMBO J 16(17):5464-5471). These proteins are particularly intriguing targets for therapeutic regulation. Within this group, TWIK-1, TREK-1 and TASK-1 and TASK-2 are widely distributed in many different tissues, while TRAAK is present exclusively in brain, spinal cord and The 4T/2P channels have different physiologic properties; TREK-1 retina. channels, are outwardly rectifying (Fink et al. (1996) EMBO J 15(24):6854-62), while TWIK-1 channels, are inwardly rectifying (Lesage et al. (1996) EMBO J 15(5):1004-11. TASK channels are regulated by changes in PH while TRAAK channels are stimulated by arachidonic acid (Reyes et al. (1998) JBC **273**(47):30863-30869).

The degree of sequence homology between different K* channel gen s is substantial. At the amino acid level, there is about 40% similarity between

different human genes, with distinct regions having higher homology, specifically the pore domain. It has been estimated that the K+ channel gene family contains approximately 10²-10³ individual genes. Despite the large number of potential genes, an analysis of public sequence databases and the scientific literature demonstrates that only a small number, approximately 20-30, have been identified. This analysis suggests that many of these important genes remain to be identified.

Potassium channels are involved in multiple different processes and are important regulators of homeostasis in nearly all cell types. Their relevance to basic cellular physiology and role in many human diseases suggests that pharmacological agents could be designed to specific channel subtypes and these compounds then applied to a large market (Bulman, D.E. (1997) Hum Mol Genet 6:1679-1685; Ackerman, M.J. and Clapham D.E. (1997) NEJM 336:1575-1586, Curran, M.E. (1998) Current Opinion in Biotechnology 9:565-572). The variety of therapeutic agents that modulate K+ channel activity reflects the diversity of physiological roles and importance of K+ channels in cellular function. A difficulty encountered in therapeutic use of therapeutic agents that modify K+ channel activity is that the presently available compounds tend to be non-specific and elicit both positive and negative responses, thereby reducing clinical efficacy. To facilitate development of specific compounds it is desirable to have further characterize novel K+ channels for use in *in vitro* and *in vivo* assays.

Relevant Literature

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A large body of literature exists in the general area of potassium channels. A review of the literature may be found in the series of books, "The Ion Channel Factsbook", volumes 1-4, by Edward C. Conley and William J. Brammar, Academic Press. An overview is provided of: extracellular ligand-gated ion channels (ISBN: 0121844501), intracellular ligand-gated channels (ISBN: 012184451X), Inward rectifier and intercellular channels (ISBN: 0121844528), and voltage gated channels (ISBN: 0121844536). Hille, B. (1992) "Ionic Channels of Excitabl Membranes", 2nd Ed. Sunderland MA:Sinauer Associates, also reviews potassium channels.

Jan and Jan (1997) <u>Annu. Rev. Neurosci.</u> **20**:91-123 review cloned potassium channels from eukaryotes and prokaryotes. Ackerman and Clapham (1997) <u>N. Engl. J. Med.</u> **336**:1575-1586 discuss the basic science of ion channels in connection with clinical disease. Bulman (1997) <u>Hum. Mol. Genet.</u> **6**:1679-1685 describe some phenotypic variation in ion channel disorders.

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Stephan *et al.* (1994) Neurology **44**:1915-1920 describe a pedigree segregating a myotonia with muscular hypertrophy and hyperirritability as an autosomal dominant trait (rippling muscle disease, Ricker *et al.* (1989) Arch. Neurol. 46405-408). Electromyography demonstrated that mechanical stimulation provoked electrically silent contractions. The responsible gene was localized to the distal end of the long arm of chromosome 1, in a 12-cM region near D1S235.

Type II pseudohypoaldosteronism is the designation used for a syndrome of chronic mineralocorticoid-resistant hyperkalemia with hypertension. The primary abnormality in type II PHA is thought to be a specific defect of the renal secretory mechanism for potassium, which limits the kaliuretic response to, but not the sodium and chloride reabsorptive effect of, mineralocorticoid. By analysis of linkage in families with autosomal dominant transmission, Mansfield *et al.* (1997) Nature Genet. 16:202-205 demonstrated locus heterogeneity of the trait, with linkage of the PHA2 gene to 1q31-q42 and 17p11-q21.

Sequences of four transmembrane, two pore potassium channels have been previously described. Reyes *et al.* (1998) <u>J Biol Chem</u> **273**(47):30863-30869 discloses a pH sensitive channel. As with the related TASK-1 and TRAAK channels, the outward rectification is lost at high external K+ concentration. The TRAAK channel is described by Fink *et al.* (1998) <u>EMBO J</u> **17**(12):3297-308. A cardiac two-pore channel is described in Kim *et al.* (1998) <u>Circ Res</u> **82**(4):513-8. An open rectifier potassium channel with two pore domains in tandem and having a postsynaptic density protein binding sequence at the C terminal was cloned by Leonoudakis *et al.* (1998) <u>J Neurosci</u> **18**(3):868-77.

The electrophysiological properties of Task channels are of interest, (Duprat *et al.* (1997) <u>EMBO J</u> **16**:5464-71). TASK currents are K+-selective, instantaneous and non-inactivating. They show an outward rectification when xternal [K+] is low, which is not observed for high [K+]out, suggesting a lack of

intrinsic voltage sensitivity. The absence of activation and inactivation kinetics as well as voltage independence are characteristic of conductances referred to as leak or background conductances. TASK is very sensitive to variations of extracellular pH in a narrow physiological range, a property probably essential for its physiological function, and suggests that small pH variations may serve a communication role in the nervous system.

SUMMARY OF THE INVENTION

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Isolated nucleotide compositions and sequences are provided for *K+Hnov* genes. The *K+Hnov* nucleic acid compositions find use in identifying homologous or related genes; in producing compositions that modulate the expression or function of its encoded proteins; for gene therapy; mapping functional regions of the proteins; and in studying associated physiological pathways. In addition, modulation of the gene activity *in vivo* is used for prophylactic and therapeutic purposes, such as treatment of potassium channel defects, identification of cell type based on expression, and the like.

DESCRIPTION OF THE SPECIFIC EMBODIMENTS

Nucleic acid compositions encoding *K+Hnov* polypeptides are provided. They are used in identifying homologous or related genes; in producing compositions that modulate the expression or function of the encoded proteins; for gene therapy; mapping functional regions of the proteins; and in studying associated physiological pathways. The *K+Hnov* gene products are members of the potassium channel gene family, and have high degrees of homology to known potassium channels. The encoded polypeptides may be alpha subunits, which form the functional channel, or accessory subunits that act to modulate the channel activity.

CHARACTERIZATION OF K+HNOV

The sequence data predict that the provided *K+Hnov* genes encode potassium channels. Table 1 summarizes the DNA sequences, corresponding SEQ ID NOs, chromosomal locations, and polymorphisms. The provided

sequences may encode a predicted K*channel, e.g. voltage gated, inward rectifier, etc.; or a modulatory subunit.

Electrophysiologic characterization of ion channels is an important part of understanding channel function. Full length ion channel cDNAs may be combined with proper vectors to form expression constructs of each individual channel. Functional analyses of expressed channels can be performed in heterologous systems, or by expression in mammalian cell lines. For expression analyses in heterologous systems such as *Xenopus* oocytes, synthetic mRNA is made through *in vitro* transcription of each channel construct. mRNA is then injected, singly or in combination with interacting channel subunit mRNAs, into prepared oocytes and the cells allowed to express the channel for several days. Oocytes expressing the channel of interest are then analyzed by whole cell voltage clamp and patch clamp techniques.

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To determine the properties of each channel when expressed in mammalian cells expression vectors specific to this type of analyses may be constructed and the resultant construct used to transform the target cells (for example human embryonic kidney (HEK) cells). Both stable and transiently expressing lines may be studied using whole cell voltage clamp and patch clamp techniques. Data obtained from EP studies includes, but is not limited to: current profiles elicited by depolarization and hyperpolarization, current-voltage (I-V) relationships, voltage dependence of activation, biophysical kinetics of channel activation, deactivation, and inactivation, reversal potential, ion selectivity, gating properties and sensitivity to channel antagonists and agonists.

Heterologous or mammalian cell lines expressing the novel channels can be used to characterize small molecules and drugs which interact with the channel. The same experiments can be used to assay for novel compounds which interact with the expressed channels.

In many cases the functional ion channel formed by K+Hnov polypeptides will be heteromultimers. Heteromultimers are known to form between different voltage gated, outward rectifying potassium channel α subunits, generally comprising four subunits, and frequently associated with auxiliary, β subunits. Typically such α subunits share a six-transmembrane domain structure (S1-S6),

with one highly positively charged domain (S4) and a pore region situated between S5 and S6. Examples of such subunits are K+Hnov4, K+Hnov9, and K+Hnov12. Channels are also formed by mutimerization of subunits of the two transmembrane and one pore architecture. It is predicted that two subunits of K+Hnov49 or K+Hnov59 will be required to form a functional channel.

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Heteromultimers of greatest interest are those that form between subunits expressed in the same tissues, and are a combination of subunits from the same species. In addition, the formation of multimers between the subject polypeptides and subunits that form functional channels are of particular interest. The resulting channel may have decreased or increased conductance relative to a homomultimer, and may be altered in response to beta subunits or other modulatory molecules.

Known voltage gated K+ channel α subunits include Kv1.1-1.8 (Gutman *et al.* (1993) Sem. Neurosci. 5:101-106); Kv2.1-2.2; Kv3.1-3.4; Kv4.1-4.3; Kv5.1; Kv6.1; Kv7.1; Kv8.1; Kv9.1-9.2. The subunits capable of forming ion inducing channels include all of those in the Kv1 through Kv4; and Kv7 families. The Kv5.1, Kv6.1, Kv8.1 and Kv9.1-9.2 subunits may be electrically silent, but functional in modifying the properties in heteromultimers.

TABLE 1

		_		_		_			_	_				_		
Channel Type	ATP-sensitive inward rectifying	Voltage gated K+ channel						Delayed rectifying K+ channel	Voitage gated K+ channel	Voltage gated K+ channel			modulatory subunit		modulatory subunit	4 transmembrane domain, 2 pore domain K+ channel
Chromosome Position	2q37	unknown						2p23	8q23	Xp21		,	13q14		18912	N/A
Polymorphisms	Alternative poly(A) tail: 1236, 2395	A312C	T335C	A377G	T344C	A401G	CA410-411GG (Ala/Thr)		Alternative poly(A) tail: 2304	C321T (Pro/Leu)	A375G (Glu/Gly)	C407T (Leu/Phe)	Alternative poly(A) tail: 1427	A689G (Gly/Arg)	T365A (Ile/Asn)	N/A
Protein SEQ	SEQ ID NO:2	SEQ ID NO:4						SEQ ID NO:8	SEQ ID NO:8	SEQ ID NO:10			SEQ ID NO:12		SEQ ID NO:14	SEQ ID NO:16
CDNA SEQ	SEQ ID NO:1	SEQ ID NO:3						SEQ ID NO:5	SEQ ID NO:7	SEQ ID NO:9			SEQ ID NO:11		SEQ ID NO:13	SEQ ID NO:15
Name	K+Hnov1	K+Hnov4						K+Hnov6	K+Hnov9	K+Hnov12			K+Hnov15		K+Hnov27	K+Hnov2

4T/2P channel	chr19	N/A	SEQ ID NO:83	SEQ ID NO:82	K*Hnov59
4T/2P channel; linked to the disease loci for rippling muscle disease 1 (RMD1), and type II pseudohypoaldosteronism	1941	(ATCT), repeats in the 3' UTR sequence, starting at position 2186	SEQ ID NO:81	SEQ ID NO:80	K'Hnov48
beta-subunit.	22p13	N/A	SEQ ID NO:30	SEQ ID NO:28-29	K+Hnov44
Homology to K+ channel protein of C. elegans	8q11	G1162A; T1460A; T2496A	SEQ ID NO:27	SEQ ID NO:26	K+Hnov42
Modulatory subunit	3429	4 alternative 5' splices	SEQ ID NO:25	SEQ ID NO:21-24	K+Hnov28
6 transmembrane domain, voltage gated K+ channel	12q14	C3168T	SEQ ID NO:20	SEQ ID NO:19	K+Hnov 14
Human ortholog of murine gene, 6 transmembrane dominas, voltage gated, delayed rectifier K+ channel	N/A	N/A	SEQ ID NO:18	SEQ ID NO:17	K+Hnov 11

K+HNOV NUCLEIC ACID COMPOSITIONS

As used herein, the term "K+Hnov" is generically used to refer to any one of the provided genetic sequences listed in Table 1. Where a specific K+Hnov sequence is intended, the numerical designation, e.g. K49 or K59, will be added. Nucleic acids encoding K+Hnov potassium channels may be cDNA or genomic DNA or a fragment thereof. The term "K+Hnov gene" shall be intended to mean the open reading frame encoding any of the provided K+Hnov polypeptides, introns, as well as adjacent 5' and 3' non-coding nucleotide sequences involved in the regulation of expression, up to about 20 kb beyond the coding region, but possibly further in either direction. The gene may be introduced into an appropriate vector for extrachromosomal maintenance or for integration into a host genome.

The term "cDNA" as used herein is intended to include all nucleic acids that share the arrangement of sequence elements found in native mature mRNA species, where sequence elements are exons and 3' and 5' non-coding regions. Normally mRNA species have contiguous exons, with the intervening introns, when present, removed by nuclear RNA splicing, to create a continuous open reading frame encoding a K+Hnov protein.

A genomic sequence of interest comprises the nucleic acid present between the initiation codon and the stop codon, as defined in the listed sequences, including all of the introns that are normally present in a native chromosome. It may further include the 3' and 5' untranslated regions found in the mature mRNA. It may further include specific transcriptional and translational regulatory sequences, such as promoters, enhancers, etc., including about 1 kb, but possibly more, of flanking genomic DNA at either the 5' or 3' end of the transcribed region. The genomic DNA may be isolated as a fragment of 100 kbp or smaller; and substantially free of flanking chromosomal sequence. The genomic DNA flanking the coding region, either 3' or 5', or internal regulatory sequences as sometimes found in introns, contains sequences required for proper tissue and stage specific expression.

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The sequence of the 5' flanking region may be utilized for promoter elements, including enhancer binding sites, that provide for developmental regulation in tissues where *K+Hnov* genes are expressed. The tissue specific expression is useful for determining the pattern of expression, and for providing promoters that mimic the native pattern of expression. Naturally occurring polymorphisms in the promoter regions are useful for determining natural variations in expression, particularly those that may be associated with disease.

Alternatively, mutations may be introduced into the promoter regions to determine the effect of altering expression in experimentally defined systems. Methods for the identification of specific DNA motifs involved in the binding of transcriptional factors are known in the art, e.g. sequence similarity to known binding motifs, gel retardation studies, etc. For examples, see Blackwell et al. (1995) Mol Med 1: 194-205; Mortlock et al. (1996) Genome Res. 6: 327-33; and Joulin and Richard-Foy (1995) Eur J Biochem 232: 620-626.

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The regulatory sequences may be used to identify *cis* acting sequences required for transcriptional or translational regulation of *K+Hnov* expression, especially in different tissues or stages of development, and to identify *cis* acting sequences and *trans* acting factors that regulate or mediate *K+Hnov* expression. Such transcription or translational control regions may be operably linked to a *K+Hnov* gene in order to promote expression of wild type or altered *K+Hnov* or other proteins of interest in cultured cells, or in embryonic, fetal or adult tissues, and for gene therapy.

The nucleic acid compositions of the subject invention may encode all or a part of the subject polypeptides. Double or single stranded fragments may be obtained of the DNA sequence by chemically synthesizing oligonucleotides in accordance with conventional methods, by restriction enzyme digestion, by PCR amplification, etc. For the most part, DNA fragments will be of at least 15 nt, usually at least 18 nt or 25 nt, and may be at least about 50 nt. Such small DNA fragments are useful as primers for PCR, hybridization screening probes, etc. Larger DNA fragments, i.e. greater than 100 nt are useful for production of the ncoded polypeptide. For use in amplification reactions, such as PCR, a pair of

primers will b used. The exact composition of the primer sequences is not critical to the invention, but for most applications the primers will hybridize to the subject sequence under stringent conditions, as known in the art. It is preferable to choose a pair of primers that will generate an amplification product of at least about 50 nt, preferably at least about 100 nt. Algorithms for the selection of primer sequences are generally known, and are available in commercial software packages. Amplification primers hybridize to complementary strands of DNA, and will prime towards each other.

The K+Hnov genes are isolated and obtained in substantial purity, generally as other than an intact chromosome. Usually, the DNA will be obtained substantially free of other nucleic acid sequences that do not include a K+Hnov sequence or fragment thereof, generally being at least about 50%, usually at least about 90% pure and are typically "recombinant", i.e. flanked by one or more nucleotides with which it is not normally associated on a naturally occurring chromosome.

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The DNA may also be used to identify expression of the gene in a biological specimen. The manner in which one probes cells for the presence of particular nucleotide sequences, as genomic DNA or RNA, is well established in the literature and does not require elaboration here. DNA or mRNA is isolated from a cell sample. The mRNA may be amplified by RT-PCR, using reverse transcriptase to form a complementary DNA strand, followed by polymerase chain reaction amplification using primers specific for the subject DNA sequences. Alternatively, the mRNA sample is separated by gel electrophoresis, transferred to a suitable support, e.g. nitrocellulose, nylon, etc., and then probed with a fragment of the subject DNA as a probe. Other techniques, such as oligonucleotide ligation assays, in situ hybridizations, and hybridization to DNA probes arrayed on a solid chip may also find use. Detection of mRNA hybridizing to the subject sequence is indicative of K+Hnov gene expression in the sample.

The sequence of a K+Hnov gene, including flanking promoter regions and coding regions, may be mutated in various ways known in the art to gen rate targ ted changes in promoter strength, sequence of the encoded prot in, etc.

The DNA sequence or protein product of such a mutation will usually be substantially similar to the sequences provided herein, *i.e.* will differ by at least one nucleotide or amino acid, respectively, and may differ by at least two but not more than about ten nucleotides or amino acids. The sequence changes may be substitutions, insertions or deletions. Deletions may further include larger changes, such as deletions of a domain or exon. Other modifications of interest include epitope tagging, e.g. with the FLAG system, HA, etc. For studies of subcellular localization, fusion proteins with green fluorescent proteins (GFP) may be used.

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Techniques for *in vitro* mutagenesis of cloned genes are known. Examples of protocols for site specific mutagenesis may be found in Gustin *et al.*, *Biotechniques* 14:22 (1993); Barany, *Gene* 37:111-23 (1985); Colicelli *et al.*, *Mol Gen Genet* 199:537-9 (1985); and Prentki *et al.*, *Gene* 29:303-13 (1984). Methods for site specific mutagenesis can be found in Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual*, CSH Press 1989, pp. 15.3-15.108; Weiner *et al.*, *Gene* 126:35-41 (1993); Sayers *et al.*, *Biotechniques* 13:592-6 (1992); Jones and Winistorfer, *Biotechniques* 12:528-30 (1992); Barton *et al.*, *Nucleic Acids Res* 18:7349-55 (1990); Marotti and Tomich, *Gene Anal Tech* 6:67-70 (1989); and Zhu, *Anal Biochem* 177:120-4 (1989). Such mutated genes may be used to study structure-function relationships of *K+Hnov*, or to alter properties of the protein that affect its function or regulation.

Homologs and orthologs of K+Hnov genes are identified by any of a number of methods. A fragment of the provided cDNA may be used as a hybridization probe against a cDNA library from the target organism of interest, where low stringency conditions are used. The probe may be a large fragment, or one or more short degenerate primers. Nucleic acids having sequence similarity are detected by hybridization under low stringency conditions, for example, at 50°C and 6XSSC (0.9 M sodium chloride/0.09 M sodium citrate) and remain bound when subjected to washing at 55°C in 1XSSC (0.15 M sodium chloride/0.015 M sodium citrate). Sequence identity may be determined by hybridization under stringent conditions, for example, at 50°C or higher and

0.1XSSC (15 mM sodium chloride/01.5 mM sodium citrate). Nucleic acids having a region of substantial identity to the provided K+Hnov sequences, e.g. allelic variants, genetically altered versions of the gene, etc., bind to the provided K+Hnov sequences under stringent hybridization conditions. By using probes, particularly labeled probes of DNA sequences, one can isolate homologous or related genes. The source of homologous genes may be any species, e.g. primate species, particularly human; rodents, such as rats and mice, canines, felines, bovines, ovines, equines, yeast, nematodes, etc.

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Between mammalian species, e.g. human and mouse, homologs have substantial sequence similarity, i.e. at least 75% sequence identity between nucleotide sequences, in some cases 80 or 90% sequence identity, and may be as high as 95% sequence identity between closely related species. Sequence similarity is calculated based on a reference sequence, which may be a subset of a larger sequence, such as a conserved motif, coding region, flanking region, etc. A reference sequence will usually be at least about 18 nt long, more usually at least about 30 nt long, and may extend to the complete sequence that is being compared. Algorithms for sequence analysis are known in the art, such as BLAST, described in Altschul et al. (1990), J. Mol. Biol. 215:403-10. In general, variants of the invention have a sequence identity greater than at least about 65%, preferably at least about 75%, more preferably at least about 85%, and may be greater than at least about 90% or more as determined by the Smith-Waterman homology search algorithm as implemented in MPSRCH program (Oxford Molecular). Exemplary search parameters for use with the MPSRCH program in order to identify sequences of a desired sequence identity are as follows: gap open penalty: 12; and gap extension penalty: 1.

K+HNOV POLYPEPTIDES

The subject nucleic acid sequences may be employed for producing all or portions of K+Hnov polypeptides. For expression, an expression cassette may be employed. The expression vector will provide a transcriptional and translational initiation region, which may be inducible or constitutive, where the coding region

is operably linked under the transcriptional control of the transcriptional initiation region, and a transcriptional and translational termination region. These control regions may be native to a *K+Hnov* gene, or may be derived from exogenous sources.

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The peptide may be expressed in prokaryotes or eukaryotes in accordance with conventional ways, depending upon the purpose for expression. For large scale production of the protein, a unicellular organism, such as *E. coli, B. subtilis, S. cerevisiae*, insect cells in combination with baculovirus vectors, or cells of a higher organism such as vertebrates, particularly mammals, *e.g.* COS 7 cells, may be used as the expression host cells. In some situations, it is desirable to express the *K+Hnov* gene in eukaryotic cells, where the *K+Hnov* protein will benefit from native folding and post-translational modifications. Small peptides can also be synthesized in the laboratory. Peptides that are subsets of the complete *K+Hnov* sequence may be used to identify and investigate parts of the protein important for function, or to raise antibodies directed against these regions.

Fragments of interest include the transmembrane and pore domains, the signal sequences, regions of interaction between subunits, *etc.* Such domains will usually include at least about 20 amino acids of the provided sequence, more usually at least about 50 amino acids, and may include 100 amino acids or more, up to the complete domain. Binding contacts may be comprised of noncontiguous sequences, which are brought into proximity by the tertiary structure of the protein. The sequence of such fragments may be modified through manipulation of the coding sequence, as described above. Truncations may be performed at the carboxy or amino terminus of the fragment, e.g. to determine the minimum sequence required for biological activity.

With the availability of the protein or fragments thereof in large amounts, by employing an expression host, the protein may be isolated and purified in accordance with conventional ways. A lysate may be prepared of the expression host and the lysate purified using HPLC, exclusion chromatography, gel electrophoresis, affinity chromatography, or oth r purification technique. The

purified protein will generally be at least about 80% pure, preferably at least about 90% pure, and may be up to and including 100% pure. Pure is intended to mean free of other proteins, as well as cellular debris.

The expressed K+Hnov polypeptides are useful for the production of antibodies, where short fragments provide for antibodies specific for the particular polypeptide, and larger fragments or the entire protein allow for the production of antibodies over the surface of the polypeptide. Antibodies may be raised to the wild-type or variant forms of K+Hnov. Antibodies may be raised to isolated peptides corresponding to specific domains, e.g. the pore domain and the transmembrane domain, or to the native protein.

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Antibodies are prepared in accordance with conventional ways, where the expressed polypeptide or protein is used as an immunogen, by itself or conjugated to known immunogenic carriers, e.g. KLH, pre-S HBsAg, other viral or eukaryotic proteins, or the like. Various adjuvants may be employed, with a series of injections, as appropriate. For monoclonal antibodies, after one or more booster injections, the spleen is isolated, the lymphocytes immortalized by cell fusion, and then screened for high affinity antibody binding. The immortalized cells, i.e. hybridomas, producing the desired antibodies may then be expanded. For further description, see Monoclonal Antibodies: A Laboratory Manual, Harlow and Lane eds., Cold Spring Harbor Laboratories, Cold Spring Harbor, New York, 1988. If desired, the mRNA encoding the heavy and light chains may be isolated and mutagenized by cloning in E. coli, and the heavy and light chains mixed to further enhance the affinity of the antibody. Alternatives to in vivo immunization as a method of raising antibodies include binding to phage "display" libraries, usually in conjunction with in vitro affinity maturation.

K+HNOV GENOTYPING

The subject nucleic acid and/or polypeptide compositions may be used to genotyping and other analysis for the presence of polymorphisms in the sequence, or variation in the expression of the subject genes. Genotyping may b performed to det rmine whether a particular polymorphisms is associated with

a disease state or genetic predisposition to a disease state, particularly diseases associated with defects in excitatory properties of cells, *e.g.* cardiac, muscle, renal and neural cells. Disease of interest include rippling muscle disease, and type II psuedohypoaldosteronism.

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Clinical disorders associated with K+ channel defects include long-QT syndrome; a congenital disorder affecting 1 in 10,000-15,000. Affected individuals have a prolonged QT interval in the electrocardiogram due to a delayed repolarization of the ventricle. Genetic linkage analyses identified two loci for long QT syndrome, LQT1, in 11p15.5 and LQT2, in 7q35-36. Positional cloning techniques identified the novel K+ channel KvLQT1 on chromosome 11 while candidate gene analysis identified causative mutations in the HERG K+ channel for LQT2.

The weaver mouse exhibits several abnormal neurological symptoms, including severe ataxia, loss of granule cell neurons in the cerebellum and dopaminergic cells in the substantia nigra, as well as seizures and male infertility. A G-protein-coupled K+ channel having a mutation in the conserved pore domain has been determined to cause the disease. The pancreatic-islet \(\mathcal{G}\)-cell ATP-sensitive K+ channel (KATP) is composed of two subunits, the sulfonylurea receptor (SUR) and the inward rectifier K+ channel Kir6.2. Mutations in both SUR and Kir6.2 have been identified in patients with persistent hyperinsulinemic hypoglycemia of infancy, which is caused by unregulated secretion of insulin.

Genotyping may also be performed for pharmacogenetic analysis to assess the association between an individual's genotype and that individual's ability to react to a therapeutic agent. Differences in target sensitivity can lead to toxicity or therapeutic failure. Relationships between polymorphisms in channel expression or specificity can be used to optimize therapeutic dose administration.

Genetic polymorphisms are identified in the K+Hnov gene (examples are listed in table 1), e.g. the repeat variation in the 3' UTR of K49. Nucleic acids comprising the polymorphic sequences are used to screen patients for altered reactivity and adverse side effects in response to drugs that act on K+ channels.

K+Hnov genotyping is performed by DNA or RNA sequence and/or hybridization analysis of any convenient sample from a patient, e.g. biopsy material, blood sample, scrapings from cheek, etc. A nucleic acid sample from an individual is analyzed for the presence of polymorphisms in K+Hnov, particularly those that affect the activity, responsiveness or expression of K+Hnov. Specific sequences of interest include any polymorphism that leads to changes in basal expression in one or more tissues, to changes in the modulation of K+Hnov expression, or alterations in K+Hnov specificity and/or activity.

The effect of a polymorphism in K+Hnov gene sequence on the response to a particular agent may be determined by *in vitro* or *in vivo* assays. Such assays may include monitoring during clinical trials, testing on genetically defined cell lines, etc. The response of an individual to the agent can then be predicted by determining the K+Hnov genotype with respect to the polymorphism. Where there is a differential distribution of a polymorphism by racial background, guidelines for drug administration can be generally tailored to a particular ethnic group.

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Biochemical studies may be performed to determine whether a sequence polymorphism in a *K+Hnov* coding region or control regions is associated with disease, for example the association of *K+Hnov* 9 with idiopathic generalized epilepsy. Disease associated polymorphisms may include deletion or truncation of the gene, mutations that alter expression level, that affect the electrical activity of the channel, *etc*.

A number of methods are available for analyzing nucleic acids for the presence of a specific sequence. Where large amounts of DNA are available, genomic DNA is used directly. Alternatively, the region of interest is cloned into a suitable vector and grown in sufficient quantity for analysis. The nucleic acid may be amplified by conventional techniques, such as the polymerase chain reaction (PCR), to provide sufficient amounts for analysis. The use of the polymerase chain reaction is described in Saiki *et al.* (1985) <u>Science</u> **239**:487, and a review of current techniques may be found in Sambrook *et al.* Molecular Cloning: A Laboratory Manual, CSH Press 1989, pp.14.2–14.33. Amplification may b used

to determine whether a polymorphism is present, by using a primer that is specific for the polymorphism. Alternatively, various methods are known in the art that utilize oligonucleotide ligation as a means of detecting polymorphisms, for examples see Riley *et al.* (1990) N.A.R. 18:2887-2890; and Delahunty *et al.* (1996) Am. J. Hum. Genet. 58:1239-1246.

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A detectable label may be included in an amplification reaction. Suitable labels include fluorochromes, e.g. fluorescein isothiocyanate (FITC), rhodamine, Texas Red, phycoerythrin, allophycocyanin, 6-carboxyfluorescein (6-FAM), 2',7'-dimethoxy-4',5'- dichloro-6-carboxyfluorescein (JOE), 6-carboxy-X-rhodamine (ROX), 6-carboxy-2',4',7',4,7- hexachlorofluorescein (HEX), 5-carboxyfluorescein (5-FAM) or N,N,N',N'-tetramethyl-6- carboxyrhodamine (TAMRA), radioactive labels, e.g. 32P, 35S, 3H; etc. The label may be a two stage system, where the amplified DNA is conjugated to biotin, haptens, etc. having a high affinity binding partner, e.g. avidin, specific antibodies, etc., where the binding partner is conjugated to a detectable label. The label may be conjugated to one or both of the primers. Alternatively, the pool of nucleotides used in the amplification is labeled, so as to incorporate the label into the amplification product.

The sample nucleic acid, e.g. amplified or cloned fragment, is analyzed by one of a number of methods known in the art. The nucleic acid may be sequenced by dideoxy or other methods. Hybridization with the variant sequence may also be used to determine its presence, by Southern blots, dot blots, etc. The hybridization pattern of a control and variant sequence to an array of oligonucleotide probes immobilised on a solid support, as described in U.S. 5,445,934, or in WO95/35505, may also be used as a means of detecting the presence of variant sequences. Single strand conformational polymorphism (SSCP) analysis, denaturing gradient gel electrophoresis (DGGE), mismatch cleavage detection, and heteroduplex analysis in gel matrices are used to detect conformational changes created by DNA sequence variation as alterations in electrophoretic mobility. Alternatively, where a polymorphism creates or destroys a recognition site for a restriction endonuclease (restriction fragment length polymorphism, RFLP), the sample is digested with that endonuclease, and the

products size fractionated to determine whether the fragment was digested. Fractionation is performed by gel or capillary electrophoresis, particularly acrylamide or agarose gels.

In one embodiment of the invention, an array of oligonucleotides are provided, where discrete positions on the array are complementary to one or more of the provided sequences, e.g. oligonucleotides of at least 12 nt, frequently 20 nt, or larger, and including the sequence flanking a polymorphic position in a K+Hnov sequence; coding sequences for different K+Hnov channels, panels of ion channels comprising one or more of the provided K+ channels; etc. Such an array may comprise a series of oligonucleotides, each of which can specifically hybridize to a different polymorphism. For examples of arrays, see Hacia et al. (1996) Nature Genetics 14:441-447; Lockhart et al. (1996) Nature Biotechnol. 14:1675-1680; and De Risi et al. (1996) Nature Genetics 14:457-460.

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Screening for polymorphisms in K+Hnov may be based on the functional or antigenic characteristics of the protein. Protein truncation assays are useful in detecting deletions that may affect the biological activity of the protein. Various immunoassays designed to detect polymorphisms in K+Hnov proteins may be used in screening. Where many diverse genetic mutations lead to a particular disease phenotype, functional protein assays have proven to be effective screening tools. The activity of the encoded K+Hnov protein as a potassium channel may be determined by comparison with the wild-type protein.

Antibodies specific for a K+Hnov may be used in staining or in immunoassays. Samples, as used herein, include biological fluids such as semen, blood, cerebrospinal fluid, tears, saliva, lymph, dialysis fluid and the like; organ or tissue culture derived fluids; and fluids extracted from physiological tissues. Also included in the term are derivatives and fractions of such fluids. The cells may be dissociated, in the case of solid tissues, or tissue sections may be analyzed. Alternatively a lysate of the cells may be prepared.

Diagnosis may be performed by a number of methods to determine the absence or presence or altered amounts of normal or abnormal K+Hnov polypeptides in patient cells. For exampl, detection may utilize staining of cells

or histological sections, performed in accordance with conventional methods. The antibodies of interest are added to the cell sample, and incubated for a period of time sufficient to allow binding to the epitope, usually at least about 10 minutes. The antibody may be labeled with radioisotopes, enzymes, fluorescers, chemiluminescers, or other labels for direct detection. Alternatively, a second stage antibody or reagent is used to amplify the signal. Such reagents are well known in the art. For example, the primary antibody may be conjugated to biotin, with horseradish peroxidase-conjugated avidin added as a second stage reagent. Alternatively, the secondary antibody conjugated to a flourescent compound, e.g. flourescein, rhodamine, Texas red, etc. Final detection uses a substrate that undergoes a color change in the presence of the peroxidase. The absence or presence of antibody binding may be determined by various methods, including flow cytometry of dissociated cells, microscopy, radiography, scintillation counting, etc.

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MODULATION OF GENE EXPRESSION

The K+Hnov genes, gene fragments, or the encoded protein or protein fragments are useful in gene therapy to treat disorders associated with K+Hnov defects. Expression vectors may be used to introduce the K+Hnov gene into a cell. Such vectors generally have convenient restriction sites located near the promoter sequence to provide for the insertion of nucleic acid sequences. Transcription cassettes may be prepared comprising a transcription initiation region, the target gene or fragment thereof, and a transcriptional termination region. The transcription cassettes may be introduced into a variety of vectors, e.g. plasmid; retrovirus, e.g. lentivirus; adenovirus; and the like, where the vectors are able to transiently or stably be maintained in the cells, usually for a period of at least about one day, more usually for a period of at least about several days to several weeks.

The gene or K+Hnov protein may be introduced into tissues or host cells by any number of routes, including viral infection, microinjection, or fusion of v sicles. Jet injection may also b used for intramuscular administration, as

described by Furth et al. (1992) Anal Biochem 205:365-368. The DNA may be coated onto gold microparticles, and delivered intradermally by a particle bombardment device, or "gene gun" as described in the literature (see, for example, Tang et al. (1992) Nature 356:152-154), where gold microprojectiles are coated with the K+Hnov or DNA, then bombarded into skin cells.

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Antisense molecules can be used to down-regulate expression of K+Hnov in cells. The anti-sense reagent may be antisense oligonucleotides (ODN), particularly synthetic ODN having chemical modifications from native nucleic acids, or nucleic acid constructs that express such anti-sense molecules as RNA. The antisense sequence is complementary to the mRNA of the targeted gene, and inhibits expression of the targeted gene products. Antisense molecules inhibit gene expression through various mechanisms, e.g. by reducing the amount of mRNA available for translation, through activation of RNAse H, or steric hindrance. One or a combination of antisense molecules may be administered, where a combination may comprise multiple different sequences.

Antisense molecules may be produced by expression of all or a part of the target gene sequence in an appropriate vector, where the transcriptional initiation is oriented such that an antisense strand is produced as an RNA molecule. Alternatively, the antisense molecule is a synthetic oligonucleotide. Antisense oligonucleotides will generally be at least about 7, usually at least about 12, more usually at least about 20 nucleotides in length, and not more than about 500, usually not more than about 50, more usually not more than about 35 nucleotides in length, where the length is governed by efficiency of inhibition, specificity, including absence of cross-reactivity, and the like. It has been found that short oligonucleotides, of from 7 to 8 bases in length, can be strong and selective inhibitors of gene expression (see Wagner *et al.* (1996) Nature Biotechnology 14:840-844).

A specific region or regions of the endogenous sense strand mRNA sequence is chosen to be complemented by the antisense sequence. Selection of a specific sequence for the oligonucleotide may us an empirical method, where several candidate sequences are assayed for inhibition of expression of

the target gene in an *in vitro* or animal model. A combination of sequences may also be used, where several regions of the mRNA sequence are selected for antisense complementation.

Antisense oligonucleotides may be chemically synthesized by methods known in the art (see Wagner et al. (1993) supra. and Milligan et al., supra.) Preferred oligonucleotides are chemically modified from the native phosphodiester structure, in order to increase their intracellular stability and binding affinity. A number of such modifications have been described in the literature, which alter the chemistry of the backbone, sugars or heterocyclic bases.

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Among useful changes in the backbone chemistry are phosphorothioates; phosphorodithioates, where both of the non-bridging oxygens are substituted with sulfur; phosphoroamidites; alkyl phosphotriesters and boranophosphates. Achiral 3'-S-5'-O-3'-O'-5'-S-phosphorothioate, derivatives include phosphate phosphorothioate, 3'-CH2-5'-O-phosphonate and 3'-NH-5'-O-phosphoroamidate. Peptide nucleic acids replace the entire ribose phosphodiester backbone with a peptide linkage. Sugar modifications are also used to enhance stability and affinity. The α-anomer of deoxyribose may be used, where the base is inverted with respect to the natural β-anomer. The 2'-OH of the ribose sugar may be altered to form 2'-O-methyl or 2'-O-allyl sugars, which provides resistance to degradation without comprising affinity. Modification of the heterocyclic bases Some useful substitutions include must maintain proper base pairing. deoxyuridine for deoxythymidine; 5-methyl-2'-deoxycytidine and 5-bromo-2'deoxycytidine for deoxycytidine. 5- propynyl-2'-deoxyuridine and 5-propynyl-2'deoxycytidine have been shown to increase affinity and biological activity when substituted for deoxythymidine and deoxycytidine, respectively.

As an alternative to anti-sense inhibitors, catalytic nucleic acid compounds, e.g. ribozymes, anti-sense conjugates, etc. may be used to inhibit gene expression. Ribozymes may be synthesized in vitro and administered to the patient, or may be needed on an expression v ctor, from which the ribozyme is synthesized in the targeted cell (for example, see International patent application

WO 9523225, and Beigelman et al. (1995) <u>Nucl. Acids Res</u> 23:4434-42). Examples of oligonucleotides with catalytic activity are described in WO 9506764. Conjugates of anti-sense ODN with a metal complex, *e.g.* terpyridylCu(II), capable of mediating mRNA hydrolysis are described in Bashkin *et al.* (1995) <u>Appl</u> Biochem Biotechnol 54:43-56.

GENETICALLY ALTERED CELL OR ANIMAL MODELS FOR K+HNOV FUNCTION

The subject nucleic acids can be used to generate transgenic animals or site specific gene modifications in cell lines. Transgenic animals may be made through homologous recombination, where the normal *K+Hnov* locus is altered. Alternatively, a nucleic acid construct is randomly integrated into the genome. Vectors for stable integration include plasmids, retroviruses and other animal viruses, YACs, and the like.

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The modified cells or animals are useful in the study of K+Hnov function and regulation. For example, a series of small deletions and/or substitutions may be made in the K+Hnov gene to determine the role of different transmembrane domains in forming multimeric structures, ion channels, etc. Of interest are the use of K+Hnov to construct transgenic animal models for epilepsy and other neurological defects, where expression of K+Hnov is specifically reduced or absent. Specific constructs of interest include anti-sense K+Hnov, which will block K+Hnov expression, expression of dominant negative K+Hnov mutations, etc. One may also provide for expression of the K+Hnov gene or variants thereof in cells or tissues where it is not normally expressed or at abnormal times of development.

DNA constructs for homologous recombination will comprise at least a portion of the *K+Hnov* gene with the desired genetic modification, and will include regions of homology to the target locus. DNA constructs for random integration need not include regions of homology to mediate recombination. Conveniently, markers for positive and negative selection are included. Methods for generating cells having targeted gene modifications through homologous recombination are

known in the art. For various techniques for transfecting mammalian cells, see Keown et al. (1990) Methods in Enzymology 185:527-537.

For embryonic stem (ES) cells, an ES cell line may be employed, or embryonic cells may be obtained freshly from a host, e.g. mouse, rat, guinea pig, etc. Such cells are grown on an appropriate fibroblast-feeder layer or grown in the presence of leukemia inhibiting factor (LIF). When ES or embryonic cells have been transformed, they may be used to produce transgenic animals. After transformation, the cells are plated onto a feeder layer in an appropriate medium. Cells containing the construct may be detected by employing a selective medium. After sufficient time for colonies to grow, they are picked and analyzed for the occurrence of homologous recombination or integration of the construct. Those colonies that are positive may then be used for embryo manipulation and blastocyst injection. Blastocysts are obtained from 4 to 6 week old superovulated females. The ES cells are trypsinized, and the modified cells are injected into the blastocoel of the blastocyst. After injection, the blastocysts are returned to each uterine horn of pseudopregnant females. Females are then allowed to go to term and the resulting offspring screened for the construct. By providing for a different phenotype of the blastocyst and the genetically modified cells, chimeric progeny can be readily detected.

The chimeric animals are screened for the presence of the modified gene and males and females having the modification are mated to produce homozygous progeny. If the gene alterations cause lethality at some point in development, tissues or organs can be maintained as allogeneic or congenic grafts or transplants, or in *in vitro* culture. The transgenic animals may be any non-human mammal, such as laboratory animals, domestic animals, etc. The transgenic animals may be used in functional studies, drug screening, *etc.*, *e.g.* to determine the effect of a candidate drug on Ras or related gene activation, oncogenesis, *etc.*

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TESTING OF K+HNOV FUNCTION and RESPONSES

Potassium channels such as K+Hnov polypeptides are involved in multiple biologically important processes. Pharmacological agents designed to affect only specific channel subtypes are of particular interest. Presently available compounds tend to be non-specific and elicit both positive and negative responses, thereby reducing clinical efficacy.

The subject polypeptides may be used in *in vitro* and *in vivo* models to test the specificity of novel compounds, and of analogs and derivatives of compounds known to act on potassium channels. Numerous pharmacological agents have profound affects on K+ channel activity. As examples, Sotalol (BETAPACE) is a class III antiarrhythmic drug that prolongs cardiac action potentials by inhibiting delayed rectifier K+ channels. Sulfonylurea drugs, such as Glipizide (GLUCOTROL) and Tolazamide (TOLAMIDE) function as antidiabetic drugs by blocking ATP-sensitive K+ channels present in pancreatic islet cells, thereby regulating insulin secretion. Diazoxide (HYPERSTAT IV) is an antihypertensive drug that activates ATP-sensitive K+ channels, resulting in the relaxation of vascular smooth muscle. There are several other examples of drugs that have antidiabetic, antihypertensive, or antiarrhythmic activities. A number of drugs that activate K+ channels that have been proposed as coronary vasodilators for the treatment of both vasospastic and chronic stable angina.

The availability of multiple K+ channel subunits allows *in vitro* reconstruction of functional channels, which may comprise different alpha and beta subunits. The individual components may be modified by sequence deletion, substitution, *etc.* to determine the functional role of specific domains.

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Drug screening may be performed using an *in vitro* model, a genetically altered cell or animal, or purified K+Hnov protein, either as monomers, homomultimers or hetermultimers. One can identify ligands or substrates that bind to, modulate or mimic the action of K+Hnov. Drug screening identifies agents that provide a replacement for K+Hnov function in abnormal cells. Of particular interest are screening assays for agents that have a low toxicity for human cells. A wide variety of assays may be used for this purpose, including

monitoring cellular excitation and conductance, labeled *in vitro* protein-protein binding assays, electrophoretic mobility shift assays, immunoassays for protein binding, and the like. The purified protein may also be used for determination of three-dimensional crystal structure, which can be used for modeling intermolecular interactions.

The term "agent" as used herein describes any molecule, *e.g.* protein or pharmaceutical, with the capability of altering or mimicking the physiological function of *K+Hnov* polypeptide. Generally a plurality of assay mixtures are run in parallel with different agent concentrations to obtain a differential response to the various concentrations. Typically, one of these concentrations serves as a negative control, *i.e.* at zero concentration or below the level of detection.

Candidate agents encompass numerous chemical classes, though typically they are organic molecules, preferably small organic compounds having a molecular weight of more than 50 and less than about 2,500 daltons. Candidate agents comprise functional groups necessary for structural interaction with proteins, particularly hydrogen bonding, and typically include at least an amine, carbonyl, hydroxyl or carboxyl group, preferably at least two of the functional chemical groups. The candidate agents often comprise cyclical carbon or heterocyclic structures and/or aromatic or polyaromatic structures substituted with one or more of the above functional groups. Candidate agents are also found among biomolecules including peptides, saccharides, fatty acids, steroids, purines, pyrimidines, derivatives, structural analogs or combinations thereof.

Candidate agents are obtained from a wide variety of sources including libraries of synthetic or natural compounds. For example, numerous means are available for random and directed synthesis of a wide variety of organic compounds and biomolecules, including expression of randomized oligonucleotides and oligopeptides. Alternatively, libraries of natural compounds in the form of bacterial, fungal, plant and animal extracts are available or readily produced. Additionally, natural or synthetically produced libraries and compounds are readily modified through conventional chemical, physical and biochemical mans, and may be used to produce combinatorial libraries. Known

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pharmacological agents may be subjected to directed or random chemical modifications, such as acylation, alkylation, esterification, amidification, *etc.* to produce structural analogs.

Where the screening assay is a binding assay, one or more of the molecules may be joined to a label, where the label can directly or indirectly provide a detectable signal. Various labels include radioisotopes, fluorescers, chemiluminescers, enzymes, specific binding molecules, particles, e.g. magnetic particles, and the like. Specific binding molecules include pairs, such as biotin and streptavidin, digoxin and antidigoxin etc. For the specific binding members, the complementary member would normally be labeled with a molecule that provides for detection, in accordance with known procedures.

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A variety of other reagents may be included in the screening assay. These include reagents like salts, neutral proteins, e.g. albumin, detergents, etc that are used to facilitate optimal protein-protein binding and/or reduce non-specific or background interactions. Reagents that improve the efficiency of the assay, such as protease inhibitors, nuclease inhibitors, anti-microbial agents, etc. may be used. The mixture of components are added in any order that provides for the requisite binding. Incubations are performed at any suitable temperature, typically between 4 and 40°C. Incubation periods are selected for optimum activity, but may also be optimized to facilitate rapid high-throughput screening. Typically between 0.1 and 1 hours will be sufficient.

The compounds having the desired pharmacological activity may be administered in a physiologically acceptable carrier to a host in a variety of ways, orally, topically, parenterally e.g. subcutaneously, intraperitoneally, by viral infection, intravascularly, etc. Depending upon the manner of introduction, the compounds may be formulated in a variety of ways. The concentration of therapeutically active compound in the formulation may vary from about 0.1-100 wt.%. The pharmaceutical compositions can be prepared in various forms, such as granules, tablets, pills, suppositories, capsules, suspensions, salves, lotions and the lik. Pharmaceutical grade organic or inorganic carriers and/or diluents suitable for oral and topical use can be used to make up

compositions containing the therapeutically-active compounds. Diluents known to the art include aqueous media, vegetable and animal oils and fats. Stabilizing agents, wetting and emulsifying agents, salts for varying the osmotic pressure or buffers for securing an adequate pH value, and skin penetration enhancers can be used as auxiliary agents.

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It is to be understood that this invention is not limited to the particular methodology, protocols, cell lines, animal species or genera, and reagents described, as such may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims.

As used herein the singular forms "a", "and", and "the" include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to "a cell" includes a plurality of such cells and reference to "the cell" includes reference to one or more cells and equivalents thereof known to those skilled in the art, and so forth. All technical and scientific terms used herein have the same meaning as commonly understood to one of ordinary skill in the art to which this invention belongs unless clearly indicated otherwise.

It must be noted that as used herein and in the appended claims, the singular forms "a", "and", and "the" include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to "a complex" includes a plurality of such complexes and reference to "the formulation" includes reference to one or more formulations and equivalents thereof known to those skilled in the art, and so forth.

All publications mentioned herein are incorporated herein by reference for the purpose of describing and disclosing, for example, the methods and methodologies that are described in the publications which might be used in connection with the presently described invention. The publications discussed above and throughout the text are provided solely for their disclosure prior to the filing date of the present application. Nothing herein is to b construed as an

admission that the inventors are not entitled to antedate such disclosure by virtue of prior invention.

EXPERIMENTAL

The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how to make and use the subject invention, and are not intended to limit the scope of what is regarded as the invention. Efforts have been made to ensure accuracy with respect to the numbers used (e.g. amounts, temperature, concentrations, etc.) but some experimental errors and deviations should be allowed for. Unless otherwise indicated, parts are parts by weight, molecular weight is average molecular weight, temperature is in degrees centigrade; and pressure is at or near atmospheric.

15 Methods

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Two different types of sequence searches were performed. The first centered on the most highly conserved region of the K+ channel family, the pore domain. The pore is composed of 15-17 amino acids and can be divided into subfamilies based on the number of transmembrane segments present in the channel. Eleven variant peptide sequences corresponding to the pore domain were used in TBLASTN searches against the EST division of Genbank. Significant matches were identified, and classified into 2 categories: identical to known human K+ channels and related to known K+ channels. The pore sequences are shown in Table 2.

TABLE 2

SEQ ID NO	Genbank #	
49		TGGTGGGCTGTGGTGACCATGACAACTGTGGGCTATGGGGACATG
50	M60451	TGGTGGGCAGTGGTCATCATCACCACTGTGGGCTACGGGGACATG
51	L02752	TGGTGGGCAGTCGTCCATGACAACTGTAGGCTATGGAGACATG
52	M55515	TGGTGGGCAGTGGTAACCATGACAGTGGGTTACGGCGATATG
53	211585	TGGTGGGCTGTCACCATGACGACCCTGGGCTATGGAGACATG
54	U40990	TGGTGGGGGTGGTCACAGTCACCATCGGCTATGGGGACAAG
55	126643	TGGTGGGCAGTGGTCACCATGACCACGGTTGGCTATGGGGACATG
88	M96747	TGGTGGGCCGTGGTCACCATGACGACCCTGGGGCTATGGAGACATG
57.	M64676	TGGTGGGCTGTGGTCACCATGACGACACTGGGCTACGGAGACATG
58	M55514	TGGTGGGCTGTGGTGACCATGACAACTGTGGGGCTATGGGGACATG
89	X83582	TTCCTGTTCCCATTGAGACCGAAACAACCATTGGGTATGGCTTCCG
99	S78684	TTTTTATTCTCAATAGAGACAGAACCACCATTGGTTATGGCTACCG
61	U22413	TTCCTCTTCTCCATTGAGACCCAGACCATAGGCTATGGTTTCAG
62	U24056	TTCCTGTTCTCGGTGGAGACGCAGACCATCGGCTATGGGTTCCG
63	U52155	TTCCTCTTCCCTTGAATCCCAAACCACCATTGGCTATGGCTTCCG
2	D87291	TITCTCTTTTCCCTGGAATCCCAGACAACCATTGGCTATGGAGTCCG
65	D50582	TTCCTTTTCTCCATTGAGGTCCAAGTGACTATTGGCTTTGGGGGGGG
88	D50315	TTTCTCTTCTCCATTGAAGTTCAAGTTACCATTGGGTTTGGAGGGAG
87	U04270	GCGCTCTACTTCACCTTCAGCAGCCTCACCAGTGTGGGCTTCGGCAAC

The unique pore peptides sequences are shown in Table 3.

TABLE 3

SEQ ID NO	Amino acid sequence
68	WWAVVSMTTVGYGDM
69	WWAVVTMTTLGYGDM
70	WWGWTVTTIGYGDK
71	WWAVVTMTTVGYGDM
72	FLFSIEVQVTIGFGG
73	FLFSLESQTTIGYGV
74	FLFSIETETTIGYGY
75	FLFSIETQTTIGYGF
76	FLFSVETQTTIGYGF
77	FLFSLESQTTIGYGF
78	FLFSIETETTIGYGF
79	ALYFTFSSLTSVGFGN

The second set of experiments was based on a complex, reiterative process.

Annotated protein and DNA sequences were obtained from GenBank for all known K+ channels from all species. The TBLASTN and BLASTN programs were used to identify homologous ESTs, which were then analyzed using the BLASTX and BLASTN algorithms to identify ESTs which were related to K+ channels yet not identical to any known human K+ channel gene.

Novel human K+ channels were defined as those that had clear homology to known K+ channels from any species and were not present as identities or near identities to any human-derived sequences in any division of Genbank.

Isolation of full length cDNA sequence. EST clones were picked from the IMAGE consortium cDNA library and end-sequenced with vector primers. Gap closure was achieved either by primer walking or transposon sequencing. GeneTrapper (Life

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Technologies) was used to isolate larger cDNA clones according to the provided protocol. RACE was used to extend the sequences as necessary using standard protocols.

Sequences were assembled in Sequencher (Gene Codes). The presence of open reading frames was assessed as well as potential start codons. Potential polymorphisms were detected as sequence variants between multiple independent clones. Sequence homologies were detected using the BLAST algorithms.

The completed gene sequences and predicted amino acid sequences are provided as SEQ ID NO:1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21-24, 26 and 28-29. Polymorphisms, chromosome locations and family assignments are shown in Table 1.

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ESTs that had top human hits with >95% identity over 100 amino acids were discarded. This was based upon the inventors' experience that these sequences were usually identical to the starting probe sequences, with the differences due to sequence error. The remaining BLASTN and BLASTX outputs for each EST were examined manually, i.e., ESTs were removed from the analysis if the inventors determined that the variation from the known related probe sequence was a result of poor database sequence. Poor database sequence was usually identified as a number of 'N' nucleotides in the database sequence for a BLASTN search and as a base deletion or insertion in the database sequence, resulting in a peptide frameshift, for a BLASTX output. ESTs for which the highest scoring match was to non-related sequences were also discarded at this stage. The EST sequences that correspond to each clone are shown in Table 4.

Table 4

Genbank Accession#	K+Hnov	clone ID	Trace	IMAGE Plate Coordinates	Read 5'/3'
N39619	K+Hnov2	277113	yy51h05.s1	611p10	3'
N46767	K+Hnov2	277113	yy51h05.r1	611p10	5'
R19352	K+Hnov11	33144	yg24f12.r1	155024	5'
R44628	K+Hnov11	33144	yg24f12.s1	155024	3'

R35526	K+Hnov14	37299	yg64e08.r1	165015	5'
R73353	K+Hnov14	157854	yl10e04.r1	251g07	5'
AA397616	K+Hnov14	728558	zt79c08.r1	1787j15	5'
AA286692	K+Hnov28	700757	zs48h03.r1	1715d6	5'
AA150494	K+Hnov42	491748	zl08e07.s1	1170013	3'
AA156697	K+Hnov42	491748	zi08e07.r1	1170013	5'
AA191752	K+Hnov42	626699	zp82d06.r1	1522f12	5'
AA216446	K+Hnov42	626699	zp82d06.s1	1522f12	3'
AA430591	K+Hnov42	773611	zw51f10.r1	1904020	5'
AA236930	K+Hnov44	683888	zs01a05.s1	1671e9	3'
AA236968	K+Hnov44	683888	zs01a05.r1	1671e9	5'

EXAMPLE 2: CHROMOSOMAL LOCALIZATION

Two primers were designed in the 3'-untranslated regions of each gene sequence to amplify a product across the Stanford G3 radiation hybrid map, or the Whitehead GB4 panel. The PCR data were submitted for automatic two-point analysis. Mapping data were correlated with cytoband information and comparisons with the OMIM human gene map data base were made. The following primers were made:

K+Hnov1 on GB4

(SEQ ID NO:31) F: 5' TATCCACATCAATGGACAAAGC 3'
(SEQ ID NO:32) R: 5' TGCATAACTGGCTGGGTGTA 3'
Results: 1.71 cR from D2S331, Cytogenetic location of 2q37

K+Hnov2 on G3

15 F: 5' GTCAGGTGACCGAGTTCA 3' R: 5' GCTCCATCTCCAGATTCTTC 3'

Results: 0.0 cR from SHGC-1320, Cytogenetic location of 11q12

K+Hnov6 on GB4

20 (SEQ ID NO:33) F: 5' TGACATCACTGGATGAACTTGA 3'
(SEQ ID NO:34) R: 5' TGCCTGCAAAGTTTGAACAT 3'
Results: 5.23 cR from WI-5509, Cytogenetic location of 2p23

K+Hnov9 on GB4
25 (SEQ ID NO:35) F: 5' TGACATCACTGGATGAACTTGA 3' (SEQ ID NO:36) R: 5' TGCCTGCAAAGTTTGAACAT 3'

Results: 1.21 cR from AFM200VC7, Cytogenetic location of 8q23

K+Hnov11 on GB4

(SEQ ID NO:37) F: 5' ACCTGGTGGTATGGAAGCAT 3' (SEQ ID NO:38) R: 5' TTTCTCCTGGCCTCTACCC 3'

Results: 2.43 cR from WI-6756, Cytogenetic location of 8q23

K+Hnov12 on G3

(SEQ ID NO:39) F: 5' TCCCTCTTGGGTGACCTTC 3'

10 (SEQ ID NO:40) R: 5' ATCTTTGTCAGCCACCAGCT 3'

Results: 7.45 cR from SHGC-32925, Cytogenetic location of Xp21

K+Hnov14 on GB4

(SEQ ID NO:41) F: 5' AGGTGTGCTGCCATCTGCTGTTCG3'

15 (SEQ ID NO:42) R: 5' AGCCTATCCTCTCTGAGAGTCAGG

Results: 7.69 cR from WI-7107, Cytogenetic location of 12q14

K+Hnov28 on GB4

(SEQ ID NO:43) F: 5' AAGCAGAGTACTCATGATGCC 3'

20 (SEQ ID NO:44) R: 5' TCTGGTAGACAGTACAGTGG 3'

Results: 35.38 cR from WI-9695, Cytogenetic location of 3q29

K+Hnov42 on G3

(SEQ ID NO:45) F: 5' CATTTGGCTGGTCCAAGATG 3'

25 (SEQ ID NO:46) R: 5' AGTCATTGGTAGGGAGGTAC 3'

Results: 7.45 cR from SHGC-32925, Cytogenetic location of Xp21

K+Hnov44 on G3

(SEQ ID NO:47) F: 5' CATGCTTCTACAGTCCAGCC 3'

30 (SEQ ID NO:48) R: 5' GGTCCTCAGTTGCAGAAATC 3'

Results: 7.45 cR from SHGC-32925, Cytogenetic location of Xp21

Map positions for K+Hnov15 and K+Hnov27 were obtained from public databases. K+Hnov2 and K+Hnov4 have not been mapped.

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EXAMPLE 3: EXPRESSION ANALYSIS

RT-PCR was utilized to characterize the expression pattern of the novel ion channels. This approach used RNA from 30 different tissues to generate first strand cDNA. Total RNA was purchased (Clontech, Invitrogen) and used to synthesize first strand cDNA using M-MLV reverse transcriptase and the supplied buffer (Gibco-BRL). The 20 µl reaction contained 5 µg total RNA, 100 ng of random primers, 10 mM DTT.

0.5 mM each dNTP, and an RNAse inhibitor (Gibco-BRL). Identical reactions were set up without reverse transcriptase to control for DNA contamination in the RNA samples. The synthesis reaction proceeded for 1 hour at 37°C followed by 10 minutes at 95°C. These cDNAs, along with control cDNA synthesis reactions without reverse transcriptase, were diluted 1:5 and 2 µl of each sample were arrayed into 96-well trays, dried, and resuspended in PCR buffer prior to PCR amplification. The cDNAs were tested with primers with defined expression patterns to verify the presence of amplifiable cDNA from each tissue. Gene-specific primers were used to amplify the cDNAs in 20 µl PCR reactions with standard conditions, 2.5 mM MgCl₂, Taq Gold, and an appropriate annealing temperature.

This approach provides for relatively high-throughput analysis of gene expression in a large set of tissues in a cost-efficient manner and provides qualitative analysis of gene expression only. Modifications can be employed, such as the use of internal control primers, limited cycling parameters, and dilution series to convert this to a quantitative experiment.

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Spleen	*		•	٠	٠		ŀ	·	·	٠	٠	ŀ	L	ļ	١		
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Fetal Conf		+	•	·	•	•	*		·ŀ	٠	٠	٠	٠	L	ŀ	ŀ	
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Cerebo		٠	٠	+	٠	•	•	·	·	٠	•	٠	ŀ	1		<u>•</u>	
Brain.		٠	٠	٠	•	٠	ŀ	·	·	٠	÷	ŀ	ŀ	1		١	
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Autanut Hu.		K.Hoov1	K+Hoo/	W-Hood	Na Paris	NA LINE	N. P. CONTR	X+Hnov11	K+Hnov12	K+Hnov14	K+Hpov16		NACULA N	K+FEDOVZO	K+Hrov42	K+Hnov44	

A "+" indicates expression in the tissue, a "-" indicates no expression, and blank square indicates no data for that sample.

K+Hnov49 on Whitehead GB4 RH mapping panel:

Primer 1 (SEQ ID NO:5): 5' - CATAGCCATAGGTGAGGACT - 3'

Primer 2: (SEQ ID N:6) 5' - GAGAGGAAAACAGTCTGGGC - 3'

Results: Cytogenetic location 1q41, 4.6cR from framework marker D1S217

K+Hnov59 on Whitehead GB4 RH mapping panel

Primer 1 (SEQ ID NO:7): 5' - GGACATCGAACTAAGACCTG - 3'

Primer 2 (SEQ ID NO:8): 5' - TCCCATGCCATTCAGATCTG - 3'

10 Results: Cytogenetic location 19q13.2, 8.34cr from framework marker D19S425

EXPRESSION ANALYSIS OF K+HNOV49

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A probe was created from a fragment corresponding to nucleotides 50 to 1284 of SEQ ID NO:83 (K+Hnov49) and purified DNA fragment was labeled with [³²P]dCTP (Amersham) by the random primer method. Adult human Multiple Tissue Northern (MTM™) Blots (Clontech) were hybridized with the [³²P]-labeled fragment in ExpressHyb™ solution (Clontech) for four hours, washed to a final stringency of 0.1xSSC, 0.1% SDS at 65°C and subjected to autoradiography for 24 hours.

Analysis revealed that K+Hnov49 is expressed as an approximately 4.2kb mRNA. Expression levels of K+Hnov49 are high in brain and liver and low in kidney tissues. No mRNA was detectable on these Northern blots for heart, skeletal muscle, colon, thymus, spleen, small intestine, placenta, lung or peripheral blood leukocytes indicating either a very low level of expression or that it is not expressed in these tissues. Expression analysis was also carried out by RT-PCR across an extended series of tissues. The results of these analyses are shown in Table 4. Primer pairs used for amplification of K+Hnov49 and 59 are the same as those used for RH mapping as indicated above.

Table 4

	Adipose	Adrenal Gland	Bladder	Brain	Cerebellum	Cervix	Colon	Esophagus	Fetal Brain	Fetal Liver	Heart	He La Cell	Kidney	Liver	Lung	Mammary Gland	Pancreas	Placenta	Prostate	Rectum	ivary	Skeletal Muscle	Skin	Small Intestine	Spieen	Stomach	Testus	3 (Uterus	· · · · · · · · · · · · · · · · · · ·
#49	+	+	+	+	+	+	-	+	+	-	+	+	+	-	+	+	-	-	+	-	+	+	-	+	-	+	+	+		•
#59	_	_		-	_	+		+		+	+	٠.	-	+	+	+	+	_	+	+	+		-	+	- +	. +	+	+	+ +	۲

WHAT IS CLAIMED IS:

- 1. An isolated nucleic acid encoding a mammalian K+Hnov protein.
- 2. An isolated nucleic acid according to Claim 1, wherein said K+Hnov protein has the amino acid sequence of SEQ ID NO:2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 25, 27, 30, 81 or 83.
 - 3. An isolated nucleic acid according to Claim 1, wherein said K+Hnov protein has an amino acid sequence that is substantially identical to the amino acid sequence of SEQ ID NO:2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 25, 27, 30, 81 or 83.
- 4. An isolated nucleic acid according to Claim 1 wherein the nucleotide sequence of said nucleic acid is SEQ ID NO:1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21,
 22, 23, 24, 26, 28, 29, 80 or 82.
 - 5. An isolated nucleic acid that hybridizes under stringent conditions to a nucleic acid sequence of claim 4.
- 20 6. An expression cassette comprising a transcriptional initiation region functional in an expression host, a nucleic acid having a sequence of the isolated nucleic acid according to Claim 1 under the transcriptional regulation of said transcriptional initiation region, and a transcriptional termination region functional in said expression host.

7. A cell comprising an expression cassette according to Claim 6 as part of an extrachromosomal element or integrated into the genome of a host cell as a result of introduction of said expression cassette into said host cell, and the cellular progeny of said host cell.

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8. A method for producing mammalian K+Hnov protein, said method comprising:

growing a cell according to Claim 7, whereby said mammalian K+Hnov protein is expressed; and

- isolating said K+Hnov protein free of other proteins.
- 9. A purified polypeptide composition comprising at least 50 weight % of the protein present as a K+Hnov protein or a fragment thereof.
- A monoclonal antibody binding specifically to a K+Hnov protein.
 - 11. A non-human transgenic animal model for K+Hnov gene function wherein said transgenic animal comprises an introduced alteration in a K+Hnov gene.

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- 12. The animal model of claim 11, wherein said animal is heterozygous for said introduced alteration.
- 13. The animal model of claim 12, wherein said animal is homozygous 20 for said introduced alteration.
 - 14. The animal model of claim 12, wherein said introduced alteration is a knockout of endogenous K+Hnov gene expression.

SEQUENCE LISTING

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aat cga gct ttt tca att cgc ttt act gac aca gca gta gta gct cacAsn Arg Ala Phe Ser Ile Arg Phe Thr Asp Thr Ala Val Val Ala His165170	642
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                   330
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                                                                   1170
Ile His Ile Asn Gly Gln Ser Ile Asp Asn Phe Gln Ile Ser Glu Thr
                                   350
               345
gga ctg aca g aataagactt atccattttt taatgtatta aatacaccca
                                                                   1220
Gly Leu Thr
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2293

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Arg Trp Glu Lys Glu Gln Leu Thr Pro Arg Glu Thr Thr Phe Leu Glu	
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	180
	240 300
	351
ξ -	
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Ala Ser Pro Leu Ada 7255 20	
and are tac acc	447
aaa too aat gog cot gto cac att gat gtg ggo ggo cac atg tac acc Lys Ser Asn Ala Pro Val His Ile Asp Val Gly Gly His Met Tyr Thr 35	
Lys Ser Asn Ala Pro Val His 11e Apr 40	
tag aga atc gga aga	495
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Ser Ser Leu Ala IIII 200 50	- 45
ata asa cad cac	543
ctt ttt gat ggt aca gag ccc att gtt ttg gac agt etc add 555 Leu Phe Asp Gly Thr Glu Pro Ile Val Leu Asp Ser Leu Lys Gln His 65 70	
	501
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the age dad tad act	000
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Leu Alg 23	

95 100 105	
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Thr Lys Tyr Pro Giu Ser Aig 125 50 60.	
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Lys Ser Leu Ile Glu Glu Val Phe Pro Glu Ile Gly Asp val Met Gly	
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Asn Ser Val Ash Ala Gly Try 185 190 180 185 190 Phe Pro Leu Asn Gly Tyr Cys His Leu Asn Ser Val Gln Val Leu Glu 200 205	
Phe Pro Leu Ash Gly Tyl Cys 200 205	
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at and the	224
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cac ccc gtg att tac cac ctg ggc cag ctc gca ctt ctt ggt tac ttg His Pro Val Ile Tyr His Leu Gly Gln Leu Ala Leu Leu Gly Tyr Leu 125 130	560
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40 45	•
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55 50 Solve Leu Val Ala Ser Ser Phe Val Leu Leu Pro Ala Leu Val Leu 80 75 80 75 Solve Phe CVS	
Gly Leu Leu Val Ala Sel 75 75 75 65 70 Car Leu Gly Ala Val Tyr Phe Cys	
65 70 75 65 Trp Gly Leu Gln Gly Asp Cys Ser Leu Leu Gly Ala Val Tyr Phe Cys 95 90 90 90 Pro Gly Arg	•
Phe Ser Ser Leu Ser Thr Ile Gly Leu Glu Asp Leu Leu Pro Gly Arg	
Sor Leu His Pro Val Ile Tyr His Leu Gly Gin Leu Ara 200	
115 Leu Gly Tyr Leu Leu Leu Gly Leu Leu Ala Met Leu Ala Val Glu 135 140 140 140 140 140 140 140 14	
Leu Gly Tyr hed 200 135 140 130 135 140 130 Lys Phe Phe Arg 160 150 155 150 155	
Thr Phe Ser Glu Leu Pro Gin var 125 155 150 150 150 150 150 150 150 150 15	
Thr Phe Set State 150 155 156 157 158 159 159 150 150 155 170 175 170 175 170 175 170 175 170 175 175 170 175 175 170 175 175 170 175 175 175 175 175 175 175 175 175 175	
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ccccgcgcgg cgcgggggggggggggggggggggggg	
eccegegegg egegggess and the transfer of the t	342
Met Thr Gly Gin Ser 25 1 5 1 5 1 5 2 2 2 2 2 2 2 2 2 2 2 2	
gac gtg tcg gag gct aac gtc gag gac ggg gag atc cgc atc aat gtg Asp Val Ser Glu Ala Asn Val Glu Asp Gly Glu Ile Arg Ile Asn Val	
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gac gtg tcg gag gct aac gtc gag gac ggg gag atc cgc atc aat gtg Asp Val Ser Glu Ala Asn Val Glu Asp Gly Glu Ile Arg Ile Asn Val 10 10 10 Met Thr Gly Gln Ser Je Ser J	342
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gac gtg tcg gag gct aac gtc gag gac ggg gag atc cgc atc aat gtg Asp Val Ser Glu Ala Asn Val Glu Asp Gly Glu Ile Arg Ile Asn Val ggc ggc ttc aag agg agg ctg cgc tcg cac acg ctg ctg cgc ttc ccc Gly Gly Phe Lys Arg Arg Leu Arg Ser His Thr Leu Leu Arg Phe Pro 35	342
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•	
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3102

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Val Leu His Phe Tyr His Thr Gly Lys Leu His Val Met Ala Glu Leu

Cys Val Phe Ser Phe Ser Gln Glu Ile Glu Tyr Trp Gly Ile Asn Glu

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160

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Leu Pro Asp Phe Gln Ile Pro Asp Ser Gln Gly Asn Pro Gly Glu Asp

Pro Arg Phe Glu Ile Val Glu His Phe Gly Ile Ala Trp Phe Thr Phe

Glu Leu Val Ala Arg Phe Ala Val Ala Pro Asp Phe Leu Lys Phe Phe 245 250

Lys Asn Ala Leu Asn Leu Ile Asp Leu Met Ser Ile Val Pro Phe Tyr

Ile Thr Leu Val Val Asn Leu Val Val Glu Ser Thr Pro Thr Leu Ala 280 285

Asn Leu Gly Arg Val Ala Gln Val Leu Arg Leu Met Arg Ile Phe Arg 295

Ile Leu Lys Leu Ala Arg His Ser Thr Gly Leu Arg Ser Leu Gly Ala 315 Thr Leu Lys Tyr Ser Tyr Lys Glu Val Gly Leu Leu Leu Leu Tyr Leu

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Glu Glu Asn Glu Gly Leu Ala Thr Ile Pro Ala Cys Trp Trp Trp Ala 360

Thr Val Ser Met Thr Thr Val Gly Tyr Gly Asp Val Val Pro Gly Thr Thr Ala Gly Lys Leu Thr Ala Ser Ala Cys Ile Leu Ala Gly Ile Leu

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atg ctt gga gct atg ttt ggg ggg gac ttc ccc aca gct cga gac cct Met Leu Gly Ala Met Phe Gly Gly Asp Phe Pro Thr Ala Arg Asp Pro 40 45 caa ggc aat tac ttt att gat cga gat gga cct ctt ttc cga tat gtc Gln Gly Asn Tyr Phe Ile Asp Arg Asp Gly Pro Leu Phe Arg Tyr Val 65 ctc aac ttc tta aga act tca gaa ttg acc tta ccg ttg gat ttt aag Leu Asn Phe Leu Arg Thr Ser Glu Leu Thr Leu Pro Leu Asp Phe Lys 70 75	549 597
atg ctt gga gct atg ttt ggg ggg gac ttc ccc aca gct cga gac cct Met Leu Gly Ala Met Phe Gly Gly Asp Phe Pro Thr Ala Arg Asp Pro 40 45 caa ggc aat tac ttt att gat cga gat gga cct ctt ttc cga tat gtc Gln Gly Asn Tyr Phe Ile Asp Arg Asp Gly Pro Leu Phe Arg Tyr Val 55 ctc aac ttc tta aga act tca gaa ttg acc tta ccg ttg gat ttt aag Leu Asn Phe Leu Arg Thr Ser Glu Leu Thr Leu Pro Leu Asp Phe Lys	549 597

38

ttg att cag tgt ctc aat gat cct aag cct ttg tat ccc atg gat act Leu Ile Gln Cys Leu Asn Asp Pro Lys Pro Leu Tyr Pro Met Asp Thr 105 110	93
ttt gaa gaa gtt gtg gag ctg tct agt act cgg aag ctt tct aag tac 7 ttt gaa gaa gtt gtg gag ctg tct agt act cgg aag ctt tct aag tac 7 phe Glu Glu Val Val Glu Leu Ser Ser Thr Arg Lys Leu Ser Lys Tyr 125	41
	189
gtc cat tcc tta cta gaa ggc atc tca aat tat ttt acc aag tgg aat gtc cat tcc tta cta gaa ggc atc tca aat tat ttt acc aag tgg aat Val His Ser Leu Leu Glu Gly Ile Ser Asn Tyr Phe Thr Lys Trp Asn 155 160	837
aag cac atg atg gac acc aga gac tgc cag gtt tcc ttt act ttt gga Lys His Met Met Asp Thr Arg Asp Cys Gln Val Ser Phe Thr Phe Gly 170 175	
ccc tgt gat tat cac cag gaa gtt tct ctt agg gtc cac ctg atg gaa CCC tgt gat tat cac cag gaa gtt tct ctt agg gtc cac ctg atg gaa CCC tgt gat tat cac cag gaa gtt tct ctt agg gtc cac ctg atg gaa CCC tgt gat tat cac cag gaa gtt tct ctt agg gtc cac ctg atg gaa 195 196 197 198	933
tac att aca aaa caa ggt ttc acg atc cgc aac acc cgg gtg cat cac Tyr Ile Thr Lys Gln Gly Phe Thr Ile Arg Asn Thr Arg Val His His 205	981
atg agt gag cgg gcc aat gaa aac aca gtg gag cac aac tgg act ttc atg agt gag cgg gcc aat gaa aac aca gtg gag cac aac tgg act ttc Met_Ser Glu Arg Ala Asn Glu Asn Thr Val Glu His Asn Trp Thr Phe 220 225	1029
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tgt agg cta gcc cgg aag aca gac gac t gatctccgac cctgccacag	
tgt agg cta gcc cgg dus Cys Arg Leu Ala Arg Lys Thr Asp Asp Cys Arg Leu Ala Arg Lys Thr Asp Asp 235	
230 aatcacagtg	1137
gttcctggaa agactctcca ggaaatggaa gatactgatt tttttttta aatcacagtg gttcctggaa agactctcca ggaaatggaa gatactgatt tttttttta aaccagagg tgagatattt tttttctttt aaatagttgt atttatttga aggcagtgag gtatgcatgt tgagatattt tttttccttt att	1197
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tgatgccttt gagaaaaatt aaggaaaact gttccaatca tttaaaagta ggaaccaaat taagtagtat tgtaatatta aaggaaaact gttctatgca tataaatcaa ggaaccaaat actgcttttt acagttatga caactgtttc tttctatgca tataaatcaa ggaaccaaat actgtatgcc atggaaatgt ctgactagaa atatttatat tgaattccca agtgtactgt atctgtagcc atggaaatgt ctctttatgc ctggtgcagt ataattccca agtgtactgt	1737
actgcttttt acagttatga tugactagaa atatttatat tgaattetga adole atetgtagee atggaaatgt ctgactagaa atatttatat tgaattetga adole atetgtgtgeagt ataatteeea agtgtaetgt teeetgtggt agaaaactta etetttatge etggtgeagt ataatteeea agtgtaetgt teeetgtggt agaaaaaaaaa aactaataaa aaatgaaata tgaaaaaaaaa aaaaaaaaa	1797 1800
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gaagacgcat cactggagdt Met Asp Asn Gly Asp Tip Gly 772 10	
1	
act gac cca gtc aca tta aat gta ggt gga cac ttg tat aca acg tct act gac cca gtc aca tta aat gta ggt gga cac ttg tat aca acg tct act gac cca gtc aca tta aat gta ggt gga cac ttg tat aca acg tct 25	459
and see gat aca tta aat gta ggt gga tac sog Tyr Thr Thr Ser	
act gac cca gtc aca tta aat gta ggt gga cac ttg tat act to the ser thr Asp Pro Val Thr Leu Asn Val Gly Gly His Leu Tyr Thr Thr Ser	
15	507
ctc acc aca ttg acg cgt tac ccg gat tcc atg ctt gga gct atg ttt ctc acc aca ttg acg cgt tac ccg gat tcc atg ctt gga gct atg ttt ctc acc aca ttg acg cgt tac ccg gat tcc atg ctt gga gct atg ttt	50.
ctc acc aca ttg acg cgt tac ccg gut Ser Met Leu Gly Ala Met Phe	
Leu Thr Thr Leu III - 35	
30	555
ggg ggg gac ttc ccc aca gct cga gac cct caa ggc aat tac ttt att ggg ggg gac ttc ccc aca gct cga gac cct caa ggc aat tac ttt att	
ggg ggg gac ttc ccc aca gct cga gac cct caa ggc aat tab Phe Ile Gly Gly Asp Phe Pro Thr Ala Arg Asp Pro Gln Gly Asn Tyr Phe Ile 50 55	
Gly Gly Asp Pile 220 50	
45	603
gat cga gat gga cct ctt ttc cga tat gtc ctc aac tte tta aga Asp Arg Asp Gly Pro Leu Phe Arg Tyr Val Leu Asn Phe Leu Arg Thr 65	
gat cga gat 950 Pro Leu Phe Arg Tyr Val Leu Abn	
ASP ALG ASP 65	651
the mat of a CEE C99	631
tca gaa ttg acc tta ccg ttg gat ttt das Slu Phe Asp Leu Leu Arg	
car Glu Leu IIII Leu - B5	
75 80 aat	699
the tag cag att gag ccc ttg att cag tgt ctc add	
aaa gaa gca gat ttt tac cag att gag ccc ttg att cag tgt ctc aat Lys Glu Ala Asp Phe Tyr Gln Ile Glu Pro Leu Ile Gln Cys Leu Asn 105	
Lys Glu Ala Asp 110 110	
man att ata qaq	747
the pag cot ttg tat coc atg gat act ttt gad gat sol Val Glu	
gat cct aag cct ttg tat ccc atg gat act ttt gaa gaa gtt get gaa get gab get gab gab get gab	
ASP PIO 2/3 - 115	795
ctg tct agt act cgg aag ctt tct aag tac tcc aac cca gtg gct gtc	,,,,
ctg tot agt act cgg aag off tot day Tyr Ser Asn Pro Val Ala Val	
ctg tct agt act cgg aag ctt tct aag tac tcc aac cca geg get ctg tct agt act cgg aag ctt tct aag tac tcc aac cca geg get ctg tct agt act ccg aac cca geg get ctg tct agt act ccg aac cca geg get ctg tct agt act ccg aac cca geg get ctg tct agt act ccg aac cca geg get ctg tct agt act ccg aac cca geg get ctg tct agt act ccg aac cca geg get ctg tct agt act ccg aac cca geg get ctg tct agt act ccg aac cca geg get ctg tct agt act ccg aac cca geg get ctg tct agt act ccg aac cca geg get ctg tct agt act ccg aac cca geg get ctg tct agt act ccg acc cc	
125 at a gaa	843
atc ata acg caa cta acc atc acc act aag gtc cat tcc tta cta gaa atc ata acg caa cta acc atc acc act aag gtc cat tcc tta cta gaa	
atc ata acg caa cta acc atc acc act aag gtc cat tee tee Glu Ile Ile Thr Gln Leu Thr Ile Thr Thr Lys Val His Ser Leu Leu Glu 145	
Ile Ile Thr Gir Los 145	
ggc atc tca aat tat ttt acc aag tgg aat aag cac atg atg gac acc ggc atc tca aat tat ttt acc aag tgg aat aag cac atg atg gac acc	891
ggc atc tca aat tat ttt acc aag tgg aat aag cac atg ts sp Thr Gly Ile Ser Asn Tyr Phe Thr Lys Trp Asn Lys His Met Met Asp Thr 160 165	
cly the Ser Asn Tyr Phe Thr Lys Trp Asn 1/5	
	939
Lat tat cac cag	,,,,
aga gac tgc cag gtt tcc ttt act tte gsb Pro Cys Asp Tyr His Gin	
Arg Asp Cys Gin var 55	
175	987
gaa gtt tct ctt agg gtc cac ctg atg gaa tac att aca aaa caa ggt	
gaa gtt tct ctt agg gtc cac ctg atg gaa tac att aca tac Gli Gly Glu Val Ser Leu Arg Val His Leu Met Glu Tyr Ile Thr Lys Gln Gly	
Glu Val Ser Led Als 195	
and god aat	1035
ttc acg atc cgc aac acc cgg gtg cat cac atg agt gag cgg gcc aat	i
ttc acg atc cgc aac acc cgg gtg cat cac atg agt gag cgs st Phe Thr Ile Arg Asn Thr Arg Val His His Met Ser Glu Arg Ala Asn 40	
40	

215	
	083
Glu Asn Thr Val Glu M25 230 225	L133 ·
aca gac gac t gatctccgac cctgccacag gttcctggaa agactctcca Thr Asp Asp	
ggaaatggaa gatactgatt tttttttta aatcacagtg tgagatattt tttttttta aatcacagaga aagttttgtg cttccccaga ttttgttccc ttccccctga gtatgcatgt gcctgttcag agtctccaga ccttaacaga tacactttttt ataaaagaat cttaataggta tataaatcac ggttcctaac tcaactagaa ggttcctaaa tgagaaaaatc tgaggaaaaatc ctagtttcca aaccaataaa taagtagtat tgagaaaatta tttaaaagta actgctttt acagttatga atcaaatca ggaacaaaat atcggaaaactta ttttctatcac tataaaccaa tataaaccaa agaacaataa taagtagtat actgctttt acagttatga agaaaactta tttttttttt	1193 1253 1313 1373 1433 1493 1553 1613 1673 1733 1793
caactgitte testasa tgaattetga atacaaaatg teetigiggo aga ctgactagaa atatttatat tgaattecca agtgtactgt ctaccagaaa aaaaaaacaa ctctttatgc ctggtgcagt ataattecca agtgtactgt ctaccagaaa aaaaaaaaaa aactaataaa aaatgaaata tgaaaaaaaaa aaaaaaaaa	1836
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gat aat gga gac tgg ggc tat atg atg act gac cca gtc aca tta aat Asp Asn Gly Asp Trp Gly Tyr Met Met Thr Asp Pro Val Thr Leu Asn 10	347
gta ggt gga cac ttg tat aca acg tct ctc acc aca ttg acg cgt tac Val Gly Gly His Leu Tyr Thr Thr Ser Leu Thr Thr Leu Thr Arg Tyr 25 30	395
ccg gat tcc atg ctt gga gct atg ttt ggg ggg gac ttc ccc aca gct Pro Asp Ser Met Leu Gly Ala Met Phe Gly Gly Asp Phe Pro Thr Ala	443
cga gac cct caa ggc aat tac ttt att gat cga gat gga cct ctt ttc cga gac cct caa ggc aat tac ttt att gat cga gat gga cct ctt ttc Arg Asp Pro Gln Gly Asn Tyr Phe Ile Asp Arg Asp Gly Pro Leu Phe 55 60 65	491
cga tat gtc ctc aac ttc tta aga act tca gaa ttg acc tta ccg ttg cga tat gtc ctc aac ttc tta aga act tca gaa ttg acc tta ccg ttg Arg Tyr Val Leu Asn Phe Leu Arg Thr Ser Glu Leu Thr Leu Pro Leu 70 75 80	539

41

gat ttt aag gaa ttt gat ctg ctt cgg aaa gaa gca gat ttt tac cag Asp Phe Lys Glu Phe Asp Leu Leu Arg Lys Glu Ala Asp Phe Tyr Gln 85 90 95	587
	635
atg gat act ttt gaa gaa gtt gtg gag ctg tct agt act cgg aag ctt Met Asp Thr Phe Glu Glu Val Val Glu Leu Ser Ser Thr Arg Lys Leu	683
tct aag tac tcc aac cca gtg gct gtc atc ata acg caa cta acc atc tct aag tac tcc aac cca gtg gct gtc atc ata acg caa cta acc atc Ser Lys Tyr Ser Asn Pro Val Ala Val Ile Ile Thr Gln Leu Thr Ile 135 140 145	731
acc act aag gtc cat tcc tta cta gaa ggc atc tca aat tat ttt acc acc act aag gtc cat tcc tta cta gaa ggc atc tca aat tat ttt acc Thr Thr Lys Val His Ser Leu Leu Glu Gly Ile Ser Asn Tyr Phe Thr 150 155	779
aag tgg aat aag cac atg atg gac acc aga gac tgc cag gtt tcc ttt Lys Trp Asn Lys His Met Met Asp Thr Arg Asp Cys Gln Val Ser Phe 165 170 175	827
act ttt gga ccc tgt gat tat cac cag gaa gtt tct ctt agg gtc cac Thr Phe Gly Pro Cys Asp Tyr His Gln Glu Val Ser Leu Arg Val His	875
ctg atg gaa tac att aca aaa caa ggt ttc acg atc cgc aac acc cgg Leu Met Glu Tyr Ile Thr Lys Gln Gly Phe Thr Ile Arg Asn Thr Arg 200 205	923
gtg cat cac atg agt gag cgg gcc aat gaa aac aca gtg gag cac aac gtg cat cac atg agt gag cgg gcc aat gaa aac aca gtg gag cac aac Val His His Met Ser Glu Arg Ala Asn Glu Asn Thr Val Glu His Asn 215 220 225	971
tgg act ttc tgt agg cta gcc cgg aag aca gac gac t gatctccgac Trp Thr Phe Cys Arg Leu Ala Arg Lys Thr Asp Asp	1018
cctgccacag gttcctggaa agactctcca ggaaatggaa gatactgatt tttttttta aaatagatgaa gatactgatt tttttttta aaatagatgaa gatactgatt tttttttta aaatagatgaa gatactgat tttttttta aaatagatgaa gatactgagagagaccaagaaggaagttttgtg ctttggcaga ctcctccatg ttttgttccc ttccccctga ttttgttccc ttccccctga taccttattt ataaaaaaaaaa	1078 1138 1198 1258 1318 1378 1438 1498 1558 1618 1678 1738
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atg act gac cca gtc aca tta aat gta ggt gga cac ttg tat aca acg Met Thr Asp Pro Val Thr Leu Asn Val Gly His Leu Tyr Thr Thr 15 20 25	162
tot oto ace aca ttg acg ogt tac ocg gat too atg ott gga got atg tot oto ace aca ttg acg ogt tac ocg gat too atg ott gga got atg tot oto acc aca ttg acg ogt tac ocg gat too atg ott gga got atg tot oto acc aca ttg acg ogt tac ocg gat too atg ott gga got atg	210
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att gat cga gat gga cct ctt ttc cga tat gtc ctc aac ttc tta aga att gat cga gat gga cct ctt ttc cga tat gtc ctc aac ttc tta aga Ile Asp Arg Asp Gly Pro Leu Phe Arg Tyr Val Leu Asn Phe Leu Arg 65	306
act tca gaa ttg acc tta ccg ttg gat ttt aag gaa ttt gat ctg ctt Thr Ser Glu Leu Thr Leu Pro Leu Asp Phe Lys Glu Phe Asp Leu Leu 80 85	354
cgg aaa gaa gca gat ttt tac cag att gag ccc ttg att cag tgt ctc cgg aaa gaa gca gat ttt tac cag att gag ccc ttg att cag tgt ctc Arg Lys Glu Ala Asp Phe Tyr Gln Ile Glu Pro Leu Ile Gln Cys Leu 105 95	402
aat gat cct aag cct ttg tat ccc atg gat act ttt gaa gaa gtt gtg aat gat cct aag cct ttg tat ccc atg gat act ttt gaa gaa gtt gtg Asn Asp Pro Lys Pro Leu Tyr Pro Met Asp Thr Phe Glu Glu Val Val 110 110	450
gag ctg tct agt act cgg aag ctt tct aag tac tcc aac cca gtg gct Glu Leu Ser Ser Thr Arg Lys Leu Ser Lys Tyr Ser Asn Pro Val Ala 135	498
gtc atc ata acg caa cta acc atc acc act aag gtc cat tcc tta cta yal Ile Ile Thr Gln Leu Thr Ile Thr Thr Lys Val His Ser Leu Leu 145	546
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International application No.
PCT/US99/03826

			PCT/US99/03826	
PC(6) :C07H	ATION OF SUBJECT MATTER 21/04; C07K 14/705; C12N 15/09, 15/63; C12Q 1/ 3.1, 24.3; 435/7.2, 69.1, 320.1; 530/350	68	100	
cording to Intern	3.1, 24.3; 435/7.2, 69.1, 320.1; 530/550 national Patent Classification (IPC) or to both nation	nal classification	and IPC	
	ADCUPD			
nimum documen	atation searched (classification system followed by	classification sy	mbols)	
J.S. : 636/23	3.1, 24.3; 435/7.2, 69.1, 320.1; 530/350			
	rched other than minimum documentation to the exte	ent that such doc	ments are included i	n the fields searched
	se consulted during the international search (name	of data base and	, where practicable,	search terms used)
Picase See Extr	F 2Boot			
	NTS CONSIDERED TO BE RELEVANT			
		-i-a- of the col	vent nessects	Relevant to claim No.
	Citation of document, with indication, where appro-			
D DA	RTISETI, M. et al. Cloning and C	Characteization	on of a Novel	1-9
	T Downtoned Potassillii		T TOMOTHERS	
Ex	pressed in Small Intestine. PEBS Lett.	1998, Vol. 4	34, pages 1/1-	
17	6, see entire document.			
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			A Comille annex	
Further (documents are listed in the continuation of Box C.		patent family annex.	international filing date or priority
• Special	estagories of estad documents:		not in conflict with the a tiple or theory underlying	
•V• qeenne	ent defining the general state of the art which is not considered if particular relevance			the alaimed invention current be
oge series	document published on or after the international filing date		ed novel or sensot be come e document is taken alone	1001 00 th this are a
	ant which may throw doubts on priority claim(s) or which is			an altimat invention cannot be
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•O• doses	and referring to an oral disclosure, use, exhibition or other	being o	brious to a person skilled	<u> </u>
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Commissione	of Patents and Trademarks	NIRMAL'S	Timure XX	- yer
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B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

APS, MEDLINE, JAPIO, BIOSIS, SCISEARCH, WPIDS, GENEMBL, NGENSEQ 34, EST, A-GENESEQ 32, PIR 58, SWISS-PROT 35, SPTREMBL 16. search terms: potassium channel, K+hnov

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be searched, the appropriate additional search fees must be paid.

Group I, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO.2, the nucleic acid having the sequence of SEQ ID NO:1, nucleic acids hybridizing to said nucleic acids, express cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:2 and K+Hnov protein of SEQ ID NO:2.

Group II, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:4, the nucleic acid having the sequence of SEQ ID NO:3, nucleic acids hybridizing to said nucleic acids, express cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:4 and K+Hnov protein of SEQ ID NO:4.

Group III, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:6, the nucleic acid having the sequence of SEQ ID NO:5, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:6 and K+Hnov protein of SEQ ID NO:6.

Group IV, claim(a)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:8, the nucleic acid having the sequence of SEQ ID NO:7, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of

SEQ ID NO:8 and K+Haov protein of SEQ ID NO:8. Group V, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:10, the nucleic acid having the sequence of SEQ ID NO:9, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of

SEQ ID NO:10 and K+Hnov protein of SEQ ID NO:10. Group VI, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:12, the nucleic acid having the sequence of SEQ ID NO:11, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Haov protein of SEQ ID NO:12 and K+Haov protein of SEQ ID NO:12.

Group VII, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:14, the nucleic acid having the sequence of SEQ ID NO:13, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Haov protein of SEQ ID NO:14 and K+Haov protein of SEQ ID NO:14.

Group VIII, claim(s)1-9, draws to sucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:16, the aucleic said having the sequence of SEQ ID NO:15, aucleic saids hybridizing to said nucleic saids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:16 and K+Hnov protein of SEQ ID NO:16.

Group IX, claim(a)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:18, the auchie acid having the sequence of SEQ ID NO:17, auchie acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:18 and K+Hnov protein of SEQ ID NO:18.

Group X, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:20, the aucleic acid having the sequence of SEQ ID NO:19, aucleic acids hybridizing to said nucleic acids, expression cassetts comprising said nucleic acids, cell comprising said expression cassette, method for producing

K+Haov protein of SEQ ID NO:20 and K+Haov protein of SEQ ID NO:20. Group XI, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:25, the nucleic acid having the sequence of SEQ ID NO:21-25, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:25 and K+Hnov protein of SEQ ID NO:25.

Group XII, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:27, the nucleic acid having the sequence of SEQ ID NO:26, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing

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K+Hnov protein of SEQ ID NO:27 and K+Hnov protein of SEQ ID NO:27.

Group XIII, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:30, the nucleic acid having the sequence of SEQ ID NO:28-29, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:30 and K+Hnov protein of SEQ ID NO:30.

Group XIV, claim(s)1-9, drawa to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:81, the nucleic acid having the sequence of SEQ ID NO:80, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:81 and K+Hnov protein of SEQ ID NO:81.

Group XV, claim(s)1-9, drawn to nucleic acids encoding K+Hnov protein having the amino acid sequence of SEQ ID NO:83, the nucleic acid having the sequence of SEQ ID NO:82, nucleic acids hybridizing to said nucleic acids, expression cassette comprising said nucleic acids, cell comprising said expression cassette, method for producing K+Hnov protein of SEQ ID NO:83 and K+Hnov protein of SEQ ID NO:83.

Group XVI, claim(s)10, drawn to monoclonal antibody that binds to K+Hnov.

Group XVII, claim(s)11-14, drawn to non-human transgenic animal model for K+Hnov.

The inventions listed as Groups I-XVII do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: Group I is directed to nucleic acid (SEQ ID NO:1) encoding the K+Hnov protein of SEQ ID NO:2, nucleic acids hybridizing to said nucleic acid, expression cassette comprising said nucleic acid, cell comprising said cassette, method of producing the K+Hnov of SEQ ID NO:2 and the protein of SEQ ID NO:2. The special technical feature is the disclosed nucleic acid of SEQ ID NO:1 encoding the K+Hnov protein of SEQ ID NO:2. The nucleic acids, proteins, antibody and transgenic animal model of Groups II-XVII do not share the special technical feature of Group I wherein the products of said Groups are structurally and functionally different. As shown in Table 1, pages 8-9, the H+Nov proteins of SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 25, 27, 30, 81 and 83 are all structurally and functionally different, the nucleic acids encoding said proteins having different chromosome positions.

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Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
2. Claims Nos.:
2. Clams Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such as extent that no meaningful international search can be carried out, specifically:
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3. Claims Nos.:
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
Please See Extra Sheet.
1. As all required additional search fees were timely paid by the applicant, this international search report covers all searcheb claims.
2. As all seasobable claims could be searched without effort justifying an additional fee, this Authority did not invite payment.
of any additional fee.
As only some of the required additional search foce were timely paid by the applicant, this international search report cover only those claims for which fees were paid, specifically claims Nos.:
4. X No required additional search fees were timely paid by the applicant. Consequently, this international search report restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
1-9, SEQ ID NO:1 and 2
Remark on Protest The additional search fees were accompanied by the applicant's protest.
No protest accompanied the payment of additional search fees.